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Importance of the vegetation-groundwater-stream continuum to understand transformation of biogenic carbon in aquatic systems – A case study based on a pine-maize comparison in a lowland sandy watershed (Landes de Gascogne, SW France)



Loris Deirmendjian ^{a,*}, Pierre Anschutz ^a, Christian Morel ^b, Alain Mollier ^b, Laurent Augusto ^b, Denis Loustau ^b, Luiz Carlos Cotovicz Jr ^{a,c}, Damien Buquet ^a, Katixa Lajaunie ^{a,d}, Gwenaëlle Chaillou ^e, Baptiste Voltz ^{a,f}, Céline Charbonnier ^a, Dominique Poirier ^a, Gwenaël Abril ^{a,c,g}

^b UMR 1391 ISPA, INRA, Bordeaux Sciences Agro, Villenave d'Ornon, 33883, France

^c Programma de pos-graduação em Geoquímica, Universidade Federal Fluminense, Outeiro São João Batista s/n, 24020015 Niterói, RJ, Brazil

^d Aix Marseille Université, CNRS/INSU, Université de Toulon, IRD, Mediterranean Institute of Oceanography (MIO) UM 110, France

^e Département Biologie, Chimie, Géographie, Université du Québec à Rimouski, Québec, Canada

^f Univ. Lille, CNRS, Univ. Littoral Côte d'Opale, UMR 8187, LOG, Laboratoire d'Océanologie et de Géosciences, F 62 930 Wimereux, France

g Biologie des Organismes et Ecosystèmes Aquatiques (BOREA), Muséum National d'Histoire Naturelle, CNRS, IRD, UPMC, UCBN, UAG, 61 rue Buffon, 75231 Paris cedex 05, France

HIGHLIGHTS

- Forest groundwater was in anoxic conditions whereas crop groundwater was in oxic conditions.
- Leaching of soil DOC occurred mostly in forest groundwater during high flow period.
- CO₂ and CH₄ were higher in forest groundwater than in crop groundwater.
- CH₄ was higher in crop streams compared to forest streams.
- CO₂ was not different between crop streams and forest streams.

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José Virgílio Cruz

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ABSTRACT

During land-aquatic transfer, carbon (C) and inorganic nutrients (IN) are transformed in soils, groundwater, and at the groundwater-surface water interface as well as in stream channels and stream sediments. However, processes and factors controlling these transfers and transformations are not well constrained, particularly with respect to land use effect. We compared C and IN concentrations in shallow groundwater and first-order streams of a sandy lowland catchment dominated by two types of land use: pine forest and maize cropland. Contrary to forest groundwater, crop groundwater exhibited oxic conditions all-year round as a result of higher evapotranspiration and better lateral drainage that decreased the water table below the organic-rich soil horizon, prevented the leaching of soil-generated dissolved organic carbon (DOC) in groundwater, and thus limited consumption of dissolved oxygen (O₂). In crop groundwater, oxic conditions inhibited denitrification and methanogenesis resulting in high nitrate (NO₃⁻; on average

* Corresponding author at: Chemical Oceanography Unit, University of Liège, Liège, Belgium. *E-mail address*: Loris.deirmendjian@uliege.be (L Deirmendjian).

^a Laboratoire Environnements et Paléoenvironnements Océaniques et Continentaux (EPOC), CNRS, Université de Bordeaux, Allée Geoffroy Saint-Hilaire, 33615 Pessac Cedex, France

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 $1140 \pm 485 \,\mu$ mol L⁻¹) and low methane (CH₄; 40 \pm 25 nmol L⁻¹) concentrations. Conversely, anoxic conditions in forest groundwater led to lower NO $_3^-$ (25 \pm 40 μ mol L⁻¹) and higher CH₄ (1770 \pm 1830 nmol L⁻¹) concentrations. The partial pressure of carbon dioxide (pCO₂; 30,650 \pm 11,590 ppmv) in crop groundwater was significantly lower than in forest groundwater ($50,630 \pm 26,070$ ppmv), and was apparently caused by the deeper water table delaying downward diffusion of soil CO₂ to the water table. In contrast, pCO₂ was not significantly different in crop (4480 \pm 2680 ppmv) and forest (4900 \pm 4500 ppmv) streams, suggesting faster degassing in forest streams resulting from greater water turbulence. Although NO₃ concentrations indicated that denitrification occurred in riparian-forest groundwater, crop streams nevertheless exhibited important signs of spring and summer eutrophication such as the development of macrophytes. Stream eutrophication favored development of anaerobic conditions in crop stream sediments, as evidenced by increased ammonia (NH_4^+) and CH_4 in stream waters and concomitant decreased in NO₃⁻ concentrations as a result of sediment denitrification. In crop streams, dredging and erosion of streambed sediments during winter sustained high concentration of particulate organic C, NH₄⁺ and CH₄. In forest streams, dissolved iron (Fe^{2+}), NH_4^+ and CH_4 were negatively correlated with O_2 reflecting the gradual oxygenation of stream water and associated oxidations of Fe^{2+} , NH_4^+ and CH_4 . The results overall showed that forest groundwater behaved as source of CO₂ and CH₄ to streams, the intensity depending on the hydrological connectivity among soils, groundwater, and streams. CH₄ production was prevented in cropland in soils and groundwater, however crop groundwater acted as a source of CO₂ to streams (but less so than forest groundwater). Conversely, in streams, pCO₂ was not significantly affected by land use while CH₄ production was enhanced by cropland. At the catchment scale, this study found substantial biogeochemical heterogeneity in C and IN concentrations between forest and crop waters, demonstrating the importance of including the full vegetation-groundwater-stream continuum when estimating land-water fluxes of C (and nitrogen) and attempting to understand their spatial and temporal dynamics.

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1. Introduction

Despite their small surface area worldwide (Downing et al., 2012), inland waters have been recognized as key component of the global carbon (C) cycle, constituting a preferential pathway of dissolved and particulate C transport from terrestrial ecosystems to the coastal ocean (Cole et al., 2007; Meybeck, 1982; Ludwig et al., 1996a, 1996b; Meybeck, 1987). Inland waters act as significant sources of carbon dioxide (CO₂) and methane (CH₄) to the atmosphere because inland waters are generally supersaturated by CO₂ and CH₄ compared to the overlying atmosphere (Abril et al., 2014; Bastviken et al., 2011; Borges et al., 2015; Lauerwald et al., 2015; Raymond et al., 2013; Stanley et al., 2016).

Inland waters and specifically small streams are tightly connected to their catchment characteristics such as hydrology and land use, as they receive large inputs of C from land (mainly from soils and groundwater), which in turn control the stream biogeochemical processes and the water composition (Aitkenhead et al., 1999; Deirmendjian and Abril, 2018; Hotchkiss et al., 2015; Johnson et al., 2006; Jones and Mulholland, 1998; McClain et al., 2003; Polsenaere and Abril, 2012; Bodmer et al., 2016; Findlay et al., 2001; Lehrter, 2006). Groundwater discharge has been recognized as an important source of CO₂ in riverine systems, especially in small streams and headwaters (Deirmendjian and Abril, 2018; Hotchkiss et al., 2015; Johnson et al., 2008; Kokic et al., 2015; Marx et al., 2017; Raymond et al., 2013; Wallin et al., 2013). On the contrary to riverine CO₂, riverine CH₄ is likely to originate from wetlands that generally combine a strong hydrological connectivity with riverine waters and a high productivity (Abril et al., 2014; Abril and Borges, 2018). Although some studies found low CH₄ concentrations in the groundwater of Belgium (up to 1.1 μ mol L⁻¹; Borges et al., 2018; Jurado et al., 2017), other studies found high CH₄ concentrations in the groundwater of Great Britain (up to 295 µmol L⁻¹; Bell et al., 2017) and in the Appalachian basin of the USA (up to 28,000 μ mol L⁻¹; Molofsky et al., 2016). Actually, soil moisture, which controls oxic/anoxic conditions in soil, is the main determinant of terrestrial CO₂ or CH₄ production in soil. As a consequence, CH₄ emissions from soils are high under strictly anaerobic conditions in waterlogged soils whereas CO₂ emissions from soils are high under aerobic conditions in drier soils (Christensen et al., 2003; Moore and Knowles, 1989). Croplands affect water mass balance at the plot scale, especially through irrigation and extraction of groundwater, which results in declining water table in many regions worldwide (Foley et al., 2005; Gleick, 2003; Jackson et al., 2001; Postel, 1999; Rosegrant et al., 2002). Investigating spatial dynamics of CO_2 and CH_4 in groundwater in relation with land use is critical better understanding processes governing their terrestrial production and leaching to groundwater.

Croplands cover about 40% of the terrestrial ice-free surface and are often associated with degradation of both ground and surface water quality (Asner et al., 2004; Clague et al., 2015; Foley et al., 2005; Hiscock et al., 1991; Ramankutty and Foley, 1999). Intensive agriculture led to an increase of nitrate (NO_3^-) entering ground and surface water environments that has fueled aquatic primary production in surface waters and led to low CO₂ and high CH₄ concentrations, the latter being related to enhanced organic matter delivery in sediments (Borges et al., 2018; Carpenter et al., 1998; Clague et al., 2015; Crawford et al., 2016; Jordan and Weller, 1996; Smith, 2003; Zhou et al., 2017). Additionally, aquatic primary production in crop streams is enhanced as a result of low light limitation (clearing of riparian vegetation), and the excessive transport of sediment-bound organic matter and nutrients to surface waters (Bernot et al., 2010; Lamba et al., 2015; Ramos et al., 2015; Young and Huryn, 1999). Soil erosion rates in agricultural landscapes are one to two times larger than those in areas with native vegetation (Montgomery, 2007; Quinton et al., 2010). Indeed, riparian forest is usually considered stream buffer zones that attenuate stream bank erosion and NO₃⁻ inputs from croplands (Balestrini et al., 2016; Cey et al., 1999; Christensen et al., 2013; Stott, 2005; Wynn and Mostaghimi, 2006). Denitrification represents a permanent removal pathway that limits the extent and impact of NO₃⁻ contamination by transforming NO₃⁻ to inert dinitrogen (N2). However, incomplete denitrification can produced nitrous oxide (N₂O), a major anthropogenic ozone-depleting substance (Ravishankara et al., 2009). On the contrary to croplands, forests are known to export fewer nutrients by limiting runoff and leakage of nutrients (Canton et al., 2012; Onderka et al., 2010).

Land use effects on both water composition and biogeochemical processes have been studied in streams and groundwater (Barnes and Raymond, 2010, 2009; Bernot et al., 2010; Bodmer et al., 2016; Jeong, 2001; Lehrter, 2006; Masese et al., 2017; Raymond and Cole, 2003; Rodrigues et al., 2018; Salvia-Castellví et al., 2005; Vidon et al., 2008; Wilson and Xenopoulos, 2009; Young and Huryn, 1999; Zhang et al., 2018), but land use studies with simultaneous groundwater and stream sampling are more scarce (Bass et al., 2014; Borges et al., 2018; Hu et al.,

2016). The objective of this study was to understand how two contrasting types of land use (pine forest and maize cropland) affected C and inorganic nutrient (IN) concentrations in shallow groundwater and in first-order streams of a sandy lowland catchment. We hypothesized that the biogeochemical variability between crop groundwater and forest groundwater was due to agricultural practices that affect N inputs (fertilizer) and water mass balance (irrigation and drainage). We hypothesized that the biogeochemical variability between crop and forest streams originate from differential lateral export of C and IN from two distinct sources (i.e., crop groundwater and forest groundwater) because of a strong hydrological connection between groundwater and streams in the studied catchment.

2. Materials and methods

2.1. Study site

The Leyre catchment (2100 km²) is located in the southwestern part of France. This is a flat coastal plain with a mean slope lower than 0.125% and a mean altitude lower than 50 m (Jolivet et al., 2007). The lithology is relatively homogeneous and composed of sandy permeable surface layers dating from the Plio-Quaternary period (Legigan, 1979; Bertran et al., 2009, 2011). The soils are podzols characterized by a low pH (\approx 4), low nutrient availability, low cationic exchange capacity, and high organic C content that can reach 50 g per kg of soil (Augusto et al., 2010; Lundström et al., 2000). In Leyre sandy podzols, the low clay and silt content causes a low soil water retention (Augusto et al., 2010).

The study area was a vast wetland until the 19^{th} century, when a wide forest of maritime pine was sown following a landscape drainage campaign resulting from an imperial decree of Napoleon III in 1857 (Jolivet et al., 2007). Currently, the catchment is mainly occupied by C₃ pine forest (approximately 85%), with a modest proportion of C₄ maize cropland (approximately 15%) (Fig. 1; Jolivet et al., 2007). Following catastrophic forest wildfires, the maize croplands were installed

Legend

during the second half of the 20th century. Consequently, their spatial distribution was not based on soil properties, as confirmed by the similar mean values of soil texture in local croplands and forests (Augusto et al., 2010; Jolivet et al., 2003). During the maize cropping season (usually May to November), farmers irrigate the plots by pumping shallow groundwater (~1-5 m deep) almost daily to maintain adequate soil moisture status, whereas maritime pine stands are never irrigated (Govind et al., 2012). As N is not limiting for tree growth in our study region (Trichet et al., 2009), forests are never fertilized with N. Conversely, croplands generally receive two N fertilizer applications annually, a first at the beginning of May $(30-50 \text{ kg N ha}^{-1})$, and second at the beginning of June with 200–250 kg N ha⁻¹ (Canton et al., 2012; Jambert et al., 1997; Ulrich et al., 2002). Additionally, in order to maintain soil pH in the 5.5-6.0 range, local maize croplands are limed with crushed limestone $(CaCO_3)$ containing a small portion of dolomite $(CaMg(CO_3)_2)$ (10 t ha^{-1} right after forest conversion and then 0.5 t ha^{-1} an^{-1} ; Iolivet et al., 2003).

The climate is oceanic with a mean annual air temperature of 13 °C and a mean annual precipitation of 930 mm (Moreaux et al., 2011). Owing to the low slope, the low soil water retention and the high permeability of the soil (i.e., hydraulic conductivity is approximately 40 cm h^{-1} , Corbier et al., 2010), the percolation of rain water is fast (55 cm h^{-1} on average, Vernier and Castro, 2010). Consequently, surface runoff does not occur as the excess of rainfall percolates into the soil and recharges the shallow groundwater, causing the water table to rise. The sandy permeable surface layers contain a free and continuous water table that is strongly interconnected with the superficial river network. This is facilitated by a dense network of drainage ditches, initiated in the 19th century and currently maintained by forest managers in order to enhance tree regeneration and growth (Thivolle-Cazat and Najar, 2001). During the sampling period, channels of some crop streams were dredged before they began to flow again. This was done to optimize local cropland drainage and to feed croplands with IN and organic residuals found in the stream sediments. To increase soil permeability and to optimize lateral drainage in local maize



Fig. 1. Land use map of the Leyre catchment showing river network and the sampling locations of groundwaters and streams.

croplands, farmers practice subsoiling and agricultural ditches are generally deeper (2.0–2.5 m) than forest ditches (1.0 m).

2.2. Sampling strategy

We defined order 0 as groundwater and order 1 as streams and ditches either having no tributaries or being seasonally dry (from June to December during our sampling period). We selected 17 sampling stations (5 shallow groundwater and 12 first-order streams) within the Leyre catchment (Table 1; Fig. 1). The groundwater sampling stations were located in maize cropland (n = 2), pine forest (n = 2; one is the Bilos station (FR-Bil) of the ICOS Research infrastructure) and in a riparian forest adjacent to a maize cropland (n = 1; Table 1; Fig. 1). The stream sampling stations were chosen based on the different proportions of croplands in their respective catchments (Table 1; Fig. 1).

Groundwater was sampled for temperature, electrical conductivity (EC), pH, dissolved oxygen (O₂), methane (CH₄), partial pressure of CO₂ (pCO₂), total alkalinity (TA), dissolved inorganic carbon (DIC), stable isotope composition of the dissolved inorganic carbon (δ^{13} C-DIC), dissolved organic carbon (DOC), ammonia (NH₄⁺), nitrate (NO₃⁻) and dissolved iron (Fe²⁺). For groundwater, we took the precaution to renew the water in the piezometers by pumping with a submersible pump before sampling. Groundwater was then sampled once the stabilization (approximately 10 min) of groundwater temperature, pH, EC and O₂ monitored with portable probes was observed. Streams were sampled for the same parameters, plus total suspended matter (TSM), particulate organic carbon (POC) and the POC content of the TSM (POC%).

2.3. Field measurements and laboratory analyses

Groundwater and streams were sampled at approximately monthly time intervals between Jan. 2014 and Jul. 2015 (Table S1). In total, throughout the sampling period, we sampled 55 groundwaters and 137 stream waters. The pCO_2 in groundwater and streams was measured directly using an equilibrator (Frankignoulle and Borges, 2001; Polsenaere et al., 2013) following the procedure of Deirmendjian and Abril (2018).

We stored the total alkalinity (TA) samples in polypropylene bottles after filtration using a syringe equipped with glass fiber filters (GF/F; 0.7 μ m). TA was then analyzed on filtered samples by automated electro-titration on 50 mL samples with 0.1 N HCl as the titrant. The equivalence point was determined from pH between 4 and 3 with the Gran method (Gran, 1952). Precision based on replicate analyses was better than $\pm 5 \mu$ mol L⁻¹. For samples with a very low pH (<4.5), we bubbled the water with atmospheric air in order to degas the CO₂. Consequently, the initial pH increased above 5, and the TA titration was then performed (Abril et al., 2015).

We calculated DIC from pCO₂, TA, and temperature measurements using the carbonic acid dissociation constants of Millero (1979) and the CO₂ solubility from Weiss (1974), using the CO₂SYS software (Lewis et al., 1998). The δ^{13} C-DIC samples were collected using 120 mL glass serum bottles sealed with a rubber stopper and treated with 0.3 mL of HgCl₂ at 20 g L⁻¹ to avoid any microbial respiration during storage. Vials were carefully sealed such that no air remained in contact with samples and were stored in the dark to prevent photooxidation. The δ^{13} C-DIC measurements were performed with the headspace technique using an isotope ratio mass spectrometer coupled to an elemental analyzer (EA-IRMS, Micromass IsoPrime) equipped with a manual gas injection port as described in Gillikin and Bouillon (2007).

 CH_4 was also measured using a headspace technique in 60 mL glass serum bottles. The headspace was created with 10 mL of N_2 gas. We then injected 0.5 mL of the headspace in a gas chromatograph equipped with a flame ionization detector (GC-FID).

DOC samples were obtained after filtration in the field through pre-combusted GF/F (0.7 μ m). DOC filtrates were stored in precombusted Pyrex vials (25 mL), acidified with 50 μ L of 37% HCl to reach pH 2, and kept at 4 °C in the laboratory before analysis. The DOC concentrations were measured with a SHIMADZU TOC 500 analyzer (in TOC-IC mode), using a technique based on thermal oxidation after a DIC removal step (Sharp, 1993). The repeatability was better than 10 μ mol L⁻¹.

Table 1

Characteristics of groundwater and stream sampling stations, ranked in decreasing order of cropland percentage in their respective sub-catchments. ^a delimited with a geographic information system software (ArcGIS 10.5®) using an hydrological database in a polyline form (BD CARTHAGE®) and a digital elevation model (BD ALTI®, resolution of 25 m), which both have been made available by the national geographic institute of France (http://www.ign.fr/). ^b retrieved with the CORINE land cover 2006 database (EEA, 2014) using a geographic information system software (ArcGIS 10.5®). ^c C, F, R corresponding to crop, forest and riparian waters, respectively, either during high or base flow. Piezometer 1 (P1) is located in a riparian mixed pine and oak forest near a first-order stream and near a maize cropland, which where P2 is located. P2 and P3 are located in the middle of two different maize croplands of 0.6 km² and 6 km², respectively. P5 is located in an 11-years old pine plot of 0.6 km² and is part of the ICOS (name is FR-Bil) research infrastructure (http://icos-ri.eu), whereas P4 is located in another pine forest (approximately same age as P5 pine forest). The depth of piezometers (from the soil surface to the bottom of the piezometer) is 5.3 m for P1, 4.9 m for P2, 9.1 m for P3, 5 m for P4 and P5.

| Stream order | Description | Catchment area (km ²) ^a | Crop (%) ^b | Forest (%) ^b | Urban (%) ^b | During high flow ^c | During base flow ^c |
|-----------------|--|--|--------------------------|----------------------------|---------------------------|-------------------------------------|-------------------------------------|
| 1 | Ditch | 1.0 | 86.5 | 13.5 | 0.0 | С | С |
| 1 | Ditch | 1.3 | 53.8 | 46.2 | 0.0 | С | С |
| 1 | Ditch | 11.3 | 44.2 | 55.8 | 0.0 | С | С |
| 1 | Ditch | 13.4 | 42.5 | 57.5 | 0.0 | С | С |
| 1 | Stream | 57.0 | 30.7 | 69.3 | 0.0 | С | С |
| 1 | Stream | 16.8 | 7.8 | 92.2 | 0.0 | F | F |
| 1 | Ditch | 7.9 | 5.8 | 94.2 | 0.0 | F | F |
| 1 | Ditch | 2.3 | 5.2 | 94.8 | 0.0 | F | С |
| 1 | Stream | 16.0 | 4.6 | 93.8 | 1.6 | С | F |
| 1 | Stream | 34.0 | 3.8 | 96.2 | 0.0 | F | F |
| 1 | Stream | 31.0 | 2.3 | 97.7 | 0.0 | F | F |
| 1 | Headwater | 0.3 | 0.0 | 100.0 | 0.0 | F | F |
| 0 | Groundwater in a riparian forest but very near (5 m) a maize cropland (P1) | | | | | R | R |
| 0 | Groundwater in maize cropland (P2) | | | | | С | С |
| 0 | Groundwater in maize cropland (P3) | | | | | С | С |
| 0 | Groundwater in pine forest (P4) | | | | | F | F |
| 0 | Groundwater in pine forest (P5) | | | | | F | F |

The water for TSM and POC measurements was filtered through preweighed and pre-combusted GF/F glass fiber filters (0.7 μ m). The filters were dried at 60 °C and stored in the dark, and subsequently, TSM was determined by gravimetry. POC was measured using the same filter. The filters were acidified in crucibles with 2 N HCl to remove carbonates and were then dried at 60 °C to remove inorganic carbon and most of the remaining acid and water (Etcheber et al., 2007). POC content was measured by combustion (1500 °C) using a LECO CS 200 analyzer and the CO₂ formed was determined quantitatively by infrared absorption. POC in μ mol L⁻¹ and POC% were then calculated. The uncertainty was $\pm 0.05\%$ of TSM.

For IN determination, water was filtered through a 0.20 μ m cellulose acetate syringe membrane. Subsamples for Fe²⁺ were acidified with 37% HCl to prevent precipitation of iron oxide, whereas subsamples for NH₄⁺ and NO₃⁻ were not acidified but kept frozen until later analyses. Then, NH₄⁺, NO₃⁻, and Fe²⁺ were analyzed by colorimetry according to standard techniques. NH₄⁺ was analyzed following the procedure of Harwood and Kühn (1970). NO₃⁻ was analyzed by flow injection analysis following the procedure of Anderson (1979). Fe²⁺ was analyzed using the ferrozine method (Stookey, 1970). Precision was ±10% for NH₄⁺ and NO₃⁻, and was ±5% for Fe²⁺.

EC, temperature, O₂, and pH were measured using portable probes (WTW®). Before each field trip, the pH probe was calibrated using two NBS buffer solutions (4 and 7), the oxygen polarographic probe was calibrated to 100% in a humid atmosphere and the conductivity probe was calibrated using a salinity standard.

2.4. Statistical analyses

K-means clustering analysis (MacQueen, 1967) was used to classify waters either as forest-dominated or as cropland-affected (Table 1). Indeed, K-means clustering analysis allows partitioning a dataset into k groups (i.e., clusters) pre-specified by the analyst (MacQueen, 1967). Contrary to forest waters at our study site, crop waters exhibit disproportionately higher NO₃⁻ concentration as a result of N fertilizer use on maize cropland (Canton et al., 2012; De Wit et al., 2005; Jambert et al., 1997, 1994). Consequently, in the K-means clustering analysis we used NO₃⁻ concentration data as a proxy to establish a statistical distinction between forest and crop waters (Table 1). K-means clustering analysis was performed one time with the groundwater dataset (but excluding the riparian groundwater) and a second time with the first-order streams dataset. We excluded data from riparian groundwater because we have considered riparian groundwater as a cluster itself (Table 1).

Principal component analysis (PCA) was used to condense multivariate information on correlated biogeochemical parameters to a set of uncorrelated variables called principal components (further referred to as dimensions). PCA was performed one time with a dataset consisting of each measured parameter in groundwater (but excluding the riparian groundwater) and a second time with the corresponding first-order streams dataset. PCA was performed separately for groundwater and streams because particulate parameters were not present in groundwater. If PCA were not performed separately for groundwater and streams, all data from groundwater would have been removed from the analysis (indeed, if one parameter is missing for a given sampling station, the sampling station is entirely deleted from the PCA). In addition, performing the PCA separately for groundwater and streams led to information that was more robust with respect to the biogeochemical variability induced by land use, in either groundwater or streams. However, to observe whether the two groundwater (crop and forest) and two streams (crop and forest) sources could be distinguished mathematically in one PCA, we performed an additional PCA with data from both groundwater and streams that excluded particulate parameters from the analysis. All concentrations data were logtransformed prior to PCA. The PCAs showed the biogeochemical variability across forest, cropland, and hydrological seasons in either groundwater or first-order streams.

Later, non-parametric bivariate analyses (Mann-Whitney statistical tests) were used to estimate if hydrological seasons or increasing stream order significantly influenced the concentration of a biogeochemical parameter. Linear regressions were performed to model the relationships between two variables by fitting a linear equation to observed data.

K-means clustering analysis (package Stats) and PCA analysis (package FactoMineR for analysis and package factoextra for visualization; Kassambra and Mundt, 2017; Lê et al., 2008) were performed with R software version 3.1.4 (R Core Team, 2018). Mann-Whitney tests and linear regressions were performed with Graph Pad Prism version 7 software.

3. Results

3.1. Hydrology

In previous work based on the same dataset, but excluding cropland sampling stations, we identified two major hydrological seasons (Deirmendjian and Abril, 2018). One defined a high flow period as two relatively short flood events that occurred in Jan. 2014-Mar. 2014 and in Feb. 2015-Mar. 2015, whereas we defined the base flow period as two longer periods of low flow occurring in Apr. 2014–Jan. 2015 and Apr. 2015-Jul. 2015. During high flow, the average and the maximum river flows were 50 m³ s⁻¹ and 119 m³ s⁻¹, respectively. During base flow, the average and the minimum river flow were 10 m³ s⁻ and 5 $m^3 s^{-1}$, respectively. The water tables in the forest, riparian forest, and cropland exhibited similar temporal fluctuations but with a different intensity, and the forest had an overall higher water table depth than the cropland (Fig. 2). The water table in the riparian area exhibited intermediate depth between the forest and cropland sites (Fig. 2). As surface runoff was negligible in the studied sandy and flat catchment, most of the stream water likely originated from groundwater discharge.

To investigate the temporal variability of the studied biogeochemical parameters, we chose to rely on hydrological regimes (high flow and base flow periods) rather than on temperature periods (seasons). At our study site climate was oceanic (by definition very temperate) and the amplitude of the water temperature was not as high as the amplitude of the river flow. As an example, Leyre River (main stem) flow could be up to 119 m³ s⁻¹ and could be down to 5 m³ s⁻¹, whereas the highest water temperature amplitude occurred in forest streams and was 6.4–25.8 °C (Table 2). Additionally, most of the lateral export



Fig. 2. Water table depth during the sampling period (Jan. 2014–Jul. 2015) across land use in the Leyre catchment. The water table in riparian area is the water table at P1 (Table 1). The water table in crop plot is the average \pm standard deviations of water tables at P2 and P3 (Table 1). The water table in forest plot is the average \pm standard deviations of water tables at P4 and P5 (Table 1).

Table 2

Values of carbon and ancillary parameters throughout sampling period (Jan. 2014-Jul. 2015) in crop and forest continuums and in riparian groundwater. Numbers between brackets are corresponding to the sampling size. For each parameter, the table showed average \pm standard deviations and the range.

| | Crop continuum | | Forest continuum | Riparian groundwater | |
|-----------------------------------|---------------------|-----------------|---------------------|----------------------|---------------------|
| | Groundwaters (22) | Streams (59) | Groundwaters (22) | Streams (78) | Groundwaters (11) |
| рН | 4.5 ± 0.2 | 6.0 ± 0.3 | 4.5 ± 0.3 | 5.8 ± 0.5 | 4.7 ± 0.1 |
| | 4.3-5.0 | 5.4-7.0 | 3.7-4.8 | 4.2-6.9 | 4.4-4.8 |
| Temperature (°C) | 14.5 ± 1.8 | 13.6 ± 4.2 | 12.8 ± 1.8 | 12.9 ± 3.9 | 14.9 ± 2.4 |
| | 10.7-17.5 | 6.4-25.8 | 8.5-15.1 | 4.8-22.1 | 11.8-17.9 |
| EC (μ S cm ⁻¹) | 360 ± 70 | 220 ± 55 | 90 ± 10 | 115 ± 30 | 160 ± 50 |
| | 220-465 | 75-370 | 65-115 | 70-200 | 95-270 |
| NO_3^- (µmol L ⁻¹) | 1140 ± 485 | 340 ± 200 | 25 ± 40 | 75 ± 70 | 310 ± 260 |
| | 260-1785 | 10-950 | 0-120 | 0-275 | 40-860 |
| NH_4^+ (µmol L ⁻¹) | 0.4 ± 0.8 | 6.1 ± 7.0 | 4.5 ± 7.0 | 1.8 ± 1.7 | 0.4 ± 0.4 |
| | 0-3.5 | 0-40 | 0.3-30 | 0-7.8 | 0-1.5 |
| Fe^{2+} (µmol L ⁻¹) | 0.9 ± 0.4 | 5.9 ± 4.4 | 15 ± 15 | 7.9 ± 12.0 | 0.6 ± 0.5 |
| | 0.1-1.9 | 0.1-22 | 0.9 ± 56 | 0.6 ± 58 | 0.2-1.5 |
| O_2 (µmol L ⁻¹) | 220 ± 65 | 290 + 45 | 20 ± 30 | 280 ± 50 | 100 ± 70 |
| | 100-315 | 160-400 | 0-110 | 110-370 | 0-170 |
| CH_4 (nmol L^{-1}) | 40 ± 25 | 460 ± 950 | 1770 ± 1830 | 240 ± 300 | 1470 ± 1490 |
| | 15-130 | 20-4900 | 50-6700 | 20-2370 | 30-4150 |
| pCO_2 (ppmv) | $30,650 \pm 11,590$ | 4480 ± 2680 | $50,630 \pm 26,070$ | 4900 ± 4500 | $42,950 \pm 28,560$ |
| | 19,000-60,550 | 1040-14,080 | 7680-116,380 | 1000-27,200 | 17,300-103,300 |
| TA (μ mol L ⁻¹) | 90 ± 25 | 100 ± 50 | 70 ± 30 | 90 ± 50 | 70 ± 15 |
| | 35-130 | 30-300 | 30-135 | 30-280 | 45-85 |
| δ^{13} C-DIC (‰) | -19.8 ± 1.3 | -18.2 ± 3.5 | -26.7 ± 1.0 | -19.8 ± 2.8 | -25.2 ± 1.1 |
| | -22 to -17.6 | -27.6 to -11.3 | -28.8 to -24 | -27.6 to -14 | -27.9 to -23.4 |
| DIC (μ mol L ⁻¹) | 1450 ± 480 | 315 ± 135 | 2460 ± 1130 | 320 ± 210 | 1960 ± 1150 |
| | 820-2590 | 90-650 | 570-5370 | 120-1280 | 940-4480 |
| DOC (μ mol L ⁻¹) | 510 ± 150 | 605 ± 320 | 930 ± 930 | 470 ± 250 | 400 ± 100 |
| | 275-880 | 220-2290 | 310-3670 | 190-1725 | 280-620 |
| TSM (mg L^{-1}) | | 5.6 ± 8.6 | | 2.3 ± 1.7 | |
| | | 0.1-50.5 | | 0.4-8.2 | |
| POC (%) | | 28 ± 10 | | 28 ± 10 | |
| · · | | 0-50 | | 10-80 | |
| POC (μ mol L ⁻¹) | | 120 ± 180 | | 50 ± 35 | |
| | | 0-1100 | | 0-170 | |

occurred during the short periods of high flow (up to 90% for DOC, Deirmendjian et al., 2018). Thus, characterizing biogeochemical variability and biogeochemical processes in relation with land use during

this hydrological period was important. Furthermore, the seasonality induced by water temperature was to a certain extent included in the defined hydrological regimes since the high flow period was associated

Table 3

Values of carbon and ancillary parameters throughout sampling period (Jan. 2014–Jul. 2015) in different types of groundwater across hydrological seasons. Numbers between brackets are corresponding to the sampling size. For each parameter, the table showed the average \pm standard deviations and the range. We defined six groups that are Cropland_HF/Cropland_BF, Forest_HF/Forest_BF and Riparian_HF/Riparian_BF corresponding to groundwaters during high flow (HF) or base flow (BF); in either cropland, forest or riparian forest.

| | Groundwater | | | | | | |
|-----------------------------------|-------------------|---------------------|---------------------|---------------------|-------------------|---------------------|--|
| | Cropland_HF (4) | Cropland_BF (18) | Forest_HF (6) | Forest_BF (16) | Riparian_HF (2) | Riparian_BF (9) | |
| pН | 4.6 ± 0.3 | 4.5 ± 0.2 | 4.4 ± 0.3 | 4.5 ± 0.3 | 4.7 ± 0.1 | 4.6 ± 0.1 | |
| - | 4.3-4.9 | 4.3-5.0 | 4.0-4.8 | 3.7-4.8 | 4.6-4.8 | 4.4-4.8 | |
| Temperature (°C) | 12.8 ± 1.7 | 14.9 ± 1.6 | 10.8 ± 1.4 | 13.5 ± 1.4 | 12.2 ± 0.6 | 15.6 ± 2.2 | |
| | 10.7-14.5 | 11.6-17.5 | 8.5-12.2 | 10.7-15.1 | 11.8-12.6 | 12.1-17.9 | |
| EC (μ S cm ⁻¹) | 370 ± 60 | 360 ± 70 | 90 ± 15 | 90 ± 10 | 200 ± 20 | 150 ± 50 | |
| | 320-460 | 220-470 | 70-115 | 70-115 | 185-215 | 95-270 | |
| NO_3^- (µmol L ⁻¹) | 1040 ± 300 | 1160 ± 420 | 30 ± 50 | 20 ± 40 | 510 ± 20 | 260 ± 270 | |
| | 760-1320 | 260-1785 | 0-120 | 0-120 | 500-520 | 40-860 | |
| NH_4^+ (µmol L ⁻¹) | 0.5 ± 0.4 | 0.4 ± 0.9 | 3.3 ± 2.2 | 5.0 ± 8.0 | 0.3 ± 0.1 | 0.4 ± 0.5 | |
| | 0.1-1 | 0-3.5 | 1.1-7 | 0-3-30 | 0.2-0.3 | 0-1.5 | |
| Fe^{2+} (µmol L ⁻¹) | 0.8 ± 0.2 | 0.9 ± 0.4 | 10.0 ± 8.2 | 15 ± 15 | 0.4 ± 0.2 | 0.7 ± 0.5 | |
| | 0.7-1.1 | 0.1-1.9 | 2.7-25.5 | 0.9-56.6 | 0.2-0.5 | 0.2-1.5 | |
| O_2 (µmol L ⁻¹) | 250 ± 90 | 220 ± 70 | 20 ± 20 | 20 ± 30 | 170 ± 0 | 100 ± 80 | |
| | 180-310 | 100-320 | 0-40 | 0-110 | 170-170 | 0-200 | |
| CH_4 (nmol L^{-1}) | 30 ± 3 | 50 ± 25 | 480 ± 630 | 2260 ± 1900 | 1460 ± 2010 | 1470 ± 1500 | |
| | 25-30 | 16–130 | 50-1700 | 50-6700 | 40-2880 | 30-4150 | |
| pCO ₂ (ppmv) | $22,050 \pm 2000$ | $32,560 \pm 12,000$ | $28,100 \pm 11,580$ | $59,080 \pm 25,060$ | $21,530 \pm 5950$ | $47,700 \pm 29,590$ | |
| _ | 19,800-24,270 | 19,000-60,550 | 7680-39,000 | 29,685-116,400 | 17,320-25,740 | 20,600-103,300 | |
| TA (μ mol L ⁻¹) | 85 ± 2 | 92 ± 30 | 95 ± 40 | 65 ± 30 | 83 ± 2 | 60-10 | |
| 10 | 82-86 | 35–130 | 60-135 | 30-100 | 82-85 | 45-75 | |
| δ^{13} C-DIC (‰) | -20.7 ± 1.1 | -19.6 ± 1.3 | -26.6 ± 1.3 | -26.8 ± 1.0 | -26.9 ± 1.4 | -24.9 ± 0.7 | |
| | -22 to -19.7 | -21.9 to -17.6 | −27.6 to −24.0 | -28.8 to -25.3 | -27.9 to -25.9 | -25.7 to -23.4 | |
| DIC (μ mol L ⁻¹) | 1100 ± 180 | 1520 ± 490 | 1500 ± 550 | 2830 ± 1080 | 1160 ± 315 | 2140 ± 1200 | |
| | 930-1300 | 820-2590 | 570-2040 | 1650-5370 | 940-1380 | 1020-4480 | |
| DOC (μ mol L ⁻¹) | 420 ± 120 | 550 ± 140 | 2230 ± 1440 | 740 ± 380 | 310 ± 50 | 420 ± 100 | |
| | 320-590 | 340-880 | 575-3670 | 310-1720 | 275-350 | 310-620 | |

Table 4

Values of carbon and ancillary parameters throughout sampling period (Jan. 2014–Jul. 2015) in different types of streams across hydrological seasons. Numbers between brackets are corresponding to the sampling size. For each parameter, the table showed the average \pm standard deviations and the range. We defined four groups that are Cropland_HF/Cropland_BF, Forest_HF/Forest_BF corresponding to streams during high flow (HF) or base flow (BF); in either cropland-affected or forest-dominated land use.

| | First-order streams | | | | | |
|------------------------------------|--|-------------------------|----------------------------|----------------------------|--|--|
| | Cropland_HF (22) | Cropland_BF (37) | Forest_HF (23) | Forest_BF (55) | | |
| рН | $\begin{array}{c} 5.9 \pm 0.3 \\ 5.4 6.6 \end{array}$ | 6.1 ± 0.4 5.5-7.0 | 5.7 ± 0.6 4.2-6.8 | 6.1 ± 0.4 5.0-6.9 | | |
| Temperature | 10.2 ± 1.6 | 15.7 ± 3.9 | 9.0 ± 1.9 | 14.6 ± 3.3 | | |
| (°C) | 6.4-12.1 | 9.1-25.8 | 4.8-12 | 8.1-22.1 | | |
| EC (μ S cm ⁻¹) | 230 ± 50 | 220 ± 60 | 110 ± 20 | 120 ± 30 | | |
| NO= (| 145-340 | /5-3/0 | 80-150 | 70-200 | | |
| NO_3 (µmol | 420 ± 220 | $290 \pm 1/0$ | 95 ± 70 | 65 ± 70 | | |
| L ') | 180-950 | 8.5-705 | 0-275 | 0-275 | | |
| NH ₄ (μ mol | 7.0 ± 8.4 | 5.5 ± 6.0 | 1.7 ± 1.7 | 1.7 ± 1.7 | | |
| E_{0}^{2+} (umol | 67 28 | 54 ± 47 | 0.3 - 7.8 | 0-0.9 | | |
| I = 1 | 0.7 ± 5.8 16-157 | $J.4 \pm 4.7$ 0.1-22 | 3.7 ± 3.0 26-136 | 0.0 ± 14.0 0.6-57.1 | | |
| $\Omega_{\rm c}$ (umol I $^{-1}$) | 1.0-15.7 200 \pm 50 | 0.1-22 | 2.0 - 15.0 300 ± 40 | 0.0-57.1 270 \pm 60 | | |
| 0 ₂ (µ1101 L) | 190 ± 30 | 160-370 | 210 - 370 | 270 ± 00 110_360 | | |
| $(H_{1} (pmol I^{-1}))$ | 130 - 400 580 \pm 1080 | 300 ± 880 | 185 ± 100 | 110-300 270 ± 340 | | |
| | 30-4380 | 20_{-4000} | 105 ± 150 | 270 ± 340 20-2370 | | |
| $p(O_{n}(nnmy))$ | 5200 ± 2370 | 4040 ± 2790 | 40-300 4200 ± 2430 | 5200 ± 5100 | | |
| pco ₂ (ppinv) | 1040-10 740 | 1220-14080 | 1240 ± 2450 | 1010-27 200 | | |
| | 1040-10,740 | 1220-14,000 | 11.690 | 1010-27,200 | | |
| TA (μ mol L ⁻¹) | 105 ± 50 | 100 ± 50 | 70 ± 40 | 95 ± 55 | | |
| | 40-300 | 30-255 | 35-195 | 30-280 | | |
| δ^{13} C-DIC (‰) | -20.6 ± 3.9 | -16.8 ± 2.4 | -22.1 ± 2.5 | -18.9 ± 2.3 | | |
| | -27.6 to | -22.3 to | -27.6 to | -23.1 to | | |
| | -11.3 | -12.4 | -16.8 | -14.0 | | |
| DIC (μ mol L ⁻¹) | 380 ± 130 | 280 ± 120 | 300 ± 150 | 330 ± 230 | | |
| | 1000-650 | 90-600 | 150-750 | 120-1280 | | |
| DOC (μ mol L ⁻¹) | 750 ± 400 | 520 ± 230 | 540 ± 305 | 450 ± 220 | | |
| | 300-2290 | 220-1520 | 260-1725 | 190-1540 | | |
| TSM (mg L^{-1}) | 9.3 ± 11.5 | 3.1 ± 4.9 | 2.8 ± 1.7 | 2.1 ± 1.7 | | |
| | 0.9-51 | 0.1-27 | 0.5-6.6 | 0.4-8.2 | | |
| POC (%) | 26 ± 10 | 30 ± 10 | 29 ± 8 | 29 ± 10 | | |
| | 15-48 | 16-48 | 20-50 | 12-80 | | |
| POC (μ mol L ⁻¹) | 190 ± 250 | 65 ± 100 | 65 ± 40 | 40 ± 30 | | |
| | 0-1100 | 0.3-540 | 0-170 | 0.5-140 | | |

with lower water temperatures while the base flow period was associated with higher water temperatures (Tables 2–4).

3.2. K-means clustering analysis

For both hydrological seasons (i.e., high and base flow), we partitioned each sampling stations (excepting riparian groundwater), into either cropland-affected or forest-dominated waters (Table 1). Kmeans clustering analysis produced satisfactory results. Logically, groundwater located in cropland was classified as crop water whereas groundwater located in forest was classified as forest water (Table 1). Stream sampling stations having >30% of croplands in their respective catchment were always classified as crop waters (Table 1). Stream sampling stations having <8% of croplands in their respective catchment were always classified as forest waters excepting two times (Table 1). These two stream sampling stations were located a few kilometers downstream from important maize croplands. Specifically, one station was a ditch strongly vegetated during the base flow period that showed signs of N fertilizer uptake from upstream cropland and, therefore, this ditch was logically classified as a crop station during base flow (Table 1). One station was a stream that exhibited a high water flow during the high flow period, which probably increased the upstream cropland influences during this hydrological period and, therefore, this stream was logically classified as a crop station during high flow (Table 1).

Excepting one strictly forested headwater, the other sampled streams were not strictly forested or cropped (Table 1). Consequently,

3.3. Land use influence on water composition of shallow groundwater

PCA on the groundwater dataset revealed that groundwater biogeochemical variability was strongly dependent on land use (maize cropland vs. pine forest) and hydrological seasons (base flow vs. high flow) (Fig. 3a, b). The first three PCA dimensions covered 44%, 17.5% and 10.5% of the total variance within the dataset, respectively (Fig. 3a, b).

PCA dimension 1 clearly separated forest groundwater from crop groundwater based on two groups of variables negatively correlated with one another (Fig. 3a). One group of variables was characterized crop groundwater and was composed of EC, NO₃⁻, δ^{13} C-DIC, and O₂, whereas the second group of variables was characterized forest groundwater and was composed of DIC, pCO₂, CH₄, Fe²⁺, and NH₄⁺ (Fig. 3a). Indeed, we observed that the yearly average of EC ($+270 \,\mu\text{S cm}^{-1}$), NO₃⁻ $(+1115 \,\mu\text{mol } L^{-1}), \delta^{13}$ C-DIC $(+6.9\%), \text{ and } O_2 (+200 \,\mu\text{mol } L^{-1})$ were higher in crop groundwater than in forest groundwater and were significantly and positively affected by cropland cover (Table 2; Fig. 4b, c, f, j). Conversely, we observed higher DIC $(+1010 \,\mu\text{mol L}^{-1})$, pCO₂ $(+19,985 \,\mu\text{mol L}^{-1})$ ppmv), CH₄ (+1730 μ mol L⁻¹), Fe²⁺ (+14.1 μ mol L⁻¹), and NH₄⁺ $(+4.1 \ \mu mol \ L^{-1})$ in forest groundwater than in crop groundwater; these were significantly and positively affected by forest cover (Table 2; Fig. 4d, e, g, h, k). In riparian groundwater, EC, NO_3^- , $\delta^{13}C$ -DIC, O₂, DIC, pCO₂, and CH₄ exhibited intermediate values between the groundwater of forest and crop sites, whereas Fe²⁺ and NH₄⁺ were low and close to those found in crop groundwater (Table 2; Fig. 4b, c, d, e, f, g, h, j, k).

In crop groundwater, EC, NO₃⁻, δ^{13} C-DIC, and O₂ were not significantly affected by hydrological seasons (Fig. 3a, S2c, h, j, k, l). However, δ^{13} C-DIC (+1.1‰), NO₃⁻ (+120 µmol L⁻¹), and DOC (+130 µmol L⁻¹) were slightly higher (but not significantly) during base flow compared to high flow (Table 3; Fig. S2c, h, j, k, l). In forest groundwater, pCO₂, DIC, CH₄, and DOC were significantly affected by hydrological seasons (Table 3; Fig. S2g, h, k, l). DOC (+1490 µmol L⁻¹) was significantly higher during high flow, whereas pCO₂ (+30,980 ppmv), DIC (+1330 µmol L⁻¹) and CH₄ (+1780 nmol L⁻¹) were significantly higher during base flow (Table 3; Fig. S2 and S4g, h, k, l). In crop and riparian groundwater, we also observed higher pCO₂ and DIC values during base flow, but with lower intensities than in forest groundwater (Table 3; Fig. S2h, k).

3.4. Land use influence on water composition of first-order streams

Fig. 3c–d present PCA based on first-order streams data set. The first three PCA dimensions covered 28.6%, 18.5% and 13.8% of the total variance within the dataset, respectively (Fig. 3c, d).

Interestingly, the PCA based on the first-order streams dataset did not clearly separate crop streams from forest streams as it did for groundwater dataset (Fig. 3a–d). This implied lower spatial variability in streams in relation to land use in than in groundwater (Tables 2–4; Fig. 4). Nevertheless, a land use gradient was observed on PCA dimension 2 (Fig. 3c, d). PCA dimension 2 was best defined by a group of variables composed of EC, CH₄, NO₃⁻, NH₄⁺, DOC, TSM, and POC, which collectively characterized crop streams (Fig. 3c, d). On a yearly average basis, significantly higher EC (+105 µS cm⁻¹), CH₄ (+220 µmol L⁻¹), NO₃⁻ (+265 µmol L⁻¹), NH₄⁺ (+4.3 µmol L⁻¹), DOC (+135 µmol L⁻¹), TSM (+3.3 mg L⁻¹), and POC (+70 µmol L⁻¹) were observed in crop streams compared to forest streams (Table 2; Fig. 4b, c, d, g, l, m, o). High CH₄, NH₄⁺, and DOC



Fig. 3. Principal component analysis (PCA) of shallow groundwater dataset (a–b) and first-order streams dataset (c–d). We represented only the first three dimensions. Numbers between brackets are corresponding to the sampling size. The sampling size in the PCAs did not correspond exactly to the sampling size in Tables 3 and 4 because R software deletes stations from the analysis with a missing value for one parameter. In these PCAs, we used all the quantitative variables measured in this study. In each PCA, we plotted as well the individuals separated in four groups. The first group corresponds to cropland-affected samples during high flow (Crop_HF), the second group corresponds to cropland-affected samples during high flow (Forest_HF) and the fourth group corresponds to forest-dominated samples during high flow (Forest_HF). The mean value of each qualitative group has 95% chance to be within the corresponding confidence ellipse.

concentrations were characteristics of forest groundwater, but in streams, these parameters were characteristic of crop streams (Fig. 3a–d). In addition, pCO₂, DIC, δ^{13} C-DIC and O₂ were not able to separate crop streams from forest streams as they did for groundwaters (Fig. 3a–d). On a yearly average basis, no significant differences were observed between crop and forest streams for these four parameters (Fig. 3f, h, j, k).

The relatively low temporal variability between high and base flow periods observed in both crop and forest streams for the studied parameters did not allow the PCA based on stream data set to clearly separate base flow samples from high flow samples (Table 4; Figs. 3c–d, S3, S4). Nevertheless, in crop streams, pH (+0.2) and δ^{13} C-DIC (+3.8‰) were significantly higher during base flow while pCO₂ (-1160 ppmv), NO₃⁻ (-130 µmol L⁻¹) and DOC (-230 µmol L⁻¹) were significantly lower



Fig. 4. Values of carbon and ancillary parameters throughout the sampling period (Jan. 2014–Jul. 2015) in groundwater and streams across land use. Histograms represent the mean with standard deviations of a given parameter. We defined four groups that are GW_Forest/GW_Crop and LForest/Lcrop corresponding to groundwaters and streams order 1 either dominated by forests or croplands. A fifth group is GW_Riparian and corresponding to riparian groundwater. Then, based on Mann-Whitney statistical analysis, we compared GW_Crop VS L_Crop, GW_Forest VS L_Forest, GW_Crop VS GW_Forest, L_Crop VS L_Forest. Three red stars (***) indicate that data were significantly different with p < 0.05. No stars indicate that data were not significantly different (p > 0.05). (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

during the same period (Table 4; Fig. S3a, j, c, h, l, m, o). In streams, TSM and POC were significantly higher during high flow but with a higher intensity in crop streams than in forest streams (Table 4; Fig. S3m, o). Interestingly, in both forest and crop streams, POC% was not significantly affected by hydrological regime (Table 4; Fig. S3n).

3.5. Upstream-downstream distribution of biogeochemical parameters

To explore the influence of land use on water composition at the groundwater-stream continuum, we observed the upstreamdownstream (groundwater-stream) distribution of biogeochemical parameters along forest and crop continuums (Table 2; Fig. 4). Along both types of continuum, some parameters (i.e., pCO₂, TA, DIC, δ^{13} C-DIC, pH, O₂) exhibited the same upstream-downstream distribution, whereas other parameters (i.e., EC, NO₃⁻, NH₄⁺, Fe²⁺, CH₄, DOC) exhibited a different upstream-downstream distribution (Table 2; Fig. 4).

In crop and forest continuums, we observed strong spatial patterns for pCO₂, TA, DIC, δ^{13} C-DIC, and pH: pCO₂ and DIC decreased while TA remained more or less constant, and δ^{13} C-DIC and pH increased (Table 2; Fig. 4a, h, i, j, k). However, a larger decrease in pCO₂ levels in the forest continuum suggested a more intense degassing in forest streams (Table 2; Fig. 4h). We also observed an increase in O₂ in both types of continuum, which could result from stream ventilation, although with a higher intensity in forest continuum (Table 2; Fig. 4f, h).

EC decreased downstream in the crop continuum, but did not in the forest continuum where EC remained very stable and much lower than in the crop continuum (Table 2; Fig. 4b). NO_3^- decreased downstream between groundwater and streams in croplands, and in contrast, in forests NO_3^- increased downstream between groundwaters and streams (Table 2; Fig. 4c). NH_4^+ , Fe^{2+} , and CH_4 decreased in the forest continuum

but they increased in the crop continuum (Table 2; Fig. 4d, e, g). DOC significantly decreased in the forest continuum but remained stable in the crop continuum (Table 2; Fig. 4l). TSM and POC were significantly higher in crop relative to forest streams; however, similarly high POC% (28%) was observed in both types of streams (Table 2; Fig. 4m, n, o).

3.6. Biogeochemistry dynamics in the groundwater-stream continuum

PCA with groundwater and streams datasets indicated mathematically that streams were fed with two distinct sources: forest groundwater mostly characterized by high pCO₂, DIC, and CH₄ concentrations and crop groundwater mostly characterized by high NO_3^- concentrations (Fig. 5). Forest and crop streams were characterized by higher O_{2} , δ^{13} C-DIC and pH values than in groundwater (Fig. 5). In this PCA, the distinction between forest and crop streams was primarily a function of NO₃, crop streams points were moved upward along dimension 2 (Fig. 5). Throughout the sampling period, we observed a negative linear relationship ($R^2 = 0.6$, p < 0.001, n = 192) between CO₂ and O₂ for all sampled groundwater and streams (Fig. 6a). On the one side, stream samples were mostly characterized by high O₂ (mean was 290 μ mol L⁻¹) and low CO₂ (mean was 4480 ppmv), excepting some forest streams during summer that were characterized by low O_2 (down to 110 μ mol L⁻¹) and high CO₂ (up to 27,200 ppmv) (Table 2; Fig. 6a, S4f, h). On the other side, anoxic conditions associated with high CO_2 (mean was 50,630 ppmv) were characteristic of forest groundwater, whereas crop groundwater exhibited O_2 (mean was 220 µmol L⁻¹) and CO₂ (mean was 30,650 ppmv) intermediate between streams and forest groundwater (Table 2; Fig. 6a). In forest groundwater, DOC was negatively and linearly correlated with CO₂ ($R^2 = 0.4$, p < 0.001, n = 22) suggesting that part of groundwater CO₂ came from degradation



Fig. 5. Principal component analysis (PCA) of shallow groundwater and stream datasets. We represented only the first three dimensions. Numbers between brackets are corresponding to the sampling size. The sampling size in the PCA did not correspond exactly to the sampling size in Table 2 because R software deletes stations from the analysis with a missing value for one parameter. In these multivariate statistical analyses, we used all the quantitative variables measured in this study. We defined four groups that are Crop_GW/Forest_GW and Crop_stream/ Forest_stream, which are corresponding to groundwater and streams order 1, either dominated by forests or croplands. The mean value of each qualitative group has 95% chance to be within the corresponding confidence ellipse.



Fig. 6. Scatter plots of (a) CO₂ (ppmv) vs. O₂ (μ mol L⁻¹), (b) CO₂ (ppmv) vs. DOC (μ mol L⁻¹), (c) CH₄ (nmol L⁻¹) vs. O₂ (μ mol L⁻¹), (d) CH₄ vs. DOC (μ mol L⁻¹), (e) CH₄(nmol L⁻¹) vs. CO₂ (ppmv), and (f) O₂ (μ mol L⁻¹) vs. DOC (μ mol L⁻¹), in all sampled groundwater and streams.

of groundwater DOC (Fig. 6b). A comparison of CO₂ and CH₄ for all sampled groundwaters and streams showed that a large portion of the CO₂ and CH₄ in forest streams could come from forest groundwater discharge (Table 2; Fig. 6e). In crop streams, CH₄, NH₄⁺, and Fe²⁺ could not originate from crop groundwater discharge since they had much lower CH_4 , NH_4^+ , and Fe^{2+} concentrations than crop streams (Table 2; Fig. 6c, 7d, f). We observed a positive linear relationship between CH₄ and NH_4^+ ($R^2 = 0.4$, p < 0.001, n = 53) in crop streams, demonstrating that these two compounds may come from the same source (Fig. 7d). Conversely, a comparison of O_2 and NH_4^+ or Fe^{2+} in forest streams indicated that NH₄⁺ and Fe²⁺ were mostly discharged through forest groundwater (Fig. 7b, f). In forest streams, the negative linear relationship between O_2 and NH_4^+ ($R^2 = 0.1$, p-value < 0.05, n = 70), Fe^{2+} $(R^2 = 0.5, p-value < 0.001, n = 77), or CH_4 (R^2 = 0.1, p-value < 0.001, n = 77)$ n = 77) suggested oxidation of these reduced compounds in the stream water column (Fig. 6c, 7b, f). We observed a gradient of NO₃⁻ concentration, from high values to low values, between crop groundwater (mean was 1140 μ mol L⁻¹), to riparian groundwater and crop streams (310 and 340 μ mol L⁻¹, respectively), to forest streams (75 μ mol L⁻¹) and to forest groundwater (25 µmol L⁻¹) (Table 2; Fig. 7a, c, e). In crop streams, a large share of riverine NO_3^- could be discharged through crop groundwater. Conversely, NO₃⁻ concentration in forest streams could not be explained by NO₃⁻ concentration in forest groundwater (Table 2; Fig. 7a, c, e). In crop groundwater, high NO₃⁻ concentrations were associated with low CH₄ concentrations. In crop streams, high NO₃⁻ concentrations could be related to high CH₄ concentrations (Fig. 7c).

4. Discussion

4.1. Water table depth in relation to land use

At the studied catchment scale, lithology, topography, soils, and precipitation are more or less uniform (Augusto et al., 2010; Jolivet et al., 2003). At the plot scale, spatial variability of water table depth in relation to land use was thus necessarily dependent on how water outputs (drainage, evapotranspiration or groundwater storage) of the water mass balance were human-affected (Govind et al., 2012; Stella et al., 2009). Local forests are never irrigated, conversely, irrigation with extraction of groundwater (that decreases groundwater storage) in local croplands could strongly bias the water mass balance at the plot scale since about half of the water diverted for irrigation is rapidly consumed through evapotranspiration (e.g., Jackson et al., 2001). Additionally, evapotranspiration in maize croplands is typically higher than in forests owing to the larger stomatal conductance that makes the exchange of C and water between the biosphere and the atmosphere much easier (Govind et al., 2012; Stella et al., 2009). Other studies have shown that the combination of subsoiling practices (increasing soil permeability) with deep agricultural ditches in croplands also affected water mass balance at the plot scale by enhancing lateral drainage of groundwater (Evans et al., 1996; Robinson et al., 1985). From an 8 year survey of local cropland, Juste et al. (1982) showed that lateral drainage strongly affected the water mass balance at the plot scale as it represented an annual mean of 637 mm (70% of the amount of precipitation), whereas precipitation was estimated at 922 mm. At the forest plot scale, lateral



Fig. 7. Scatter plots of (a) O_2 (µmol L^{-1}) vs. NO₃⁻ (µmol L^{-1}), (b) O_2 (µmol L^{-1}) vs. NH₄⁺ (µmol L^{-1}), (c) CH₄ (µmol L^{-1}) vs. NO₃⁻ (µmol L^{-1}), (d) CH₄ (µmol L^{-1}) vs. NH₄⁺ (µmol L^{-1}), (e) DOC (µmol L^{-1}) vs. NO₃⁻ (µmol L^{-1}), and (f) O_2 (µmol L^{-1}) vs. Fe²⁺ (µmol L^{-1}), in all sampled groundwater and streams.

drainage represented an annual mean of 182 mm (20% of the amount of precipitation), whereas precipitation was estimated at 895 mm (Deirmendjian et al., 2018). At the study site, deeper water table in croplands was thus a consequence of a higher evapotranspiration and more lateral drainage than in forest. As explained further, water table depth is an important determinant for understanding the biogeochemical variability in groundwater in relation to land use.

4.2. Dynamic of O_2 , DOC, DIC and δ^{13} C-DIC in groundwater: A combination of hydrological, physical and metabolic processes

In other aquifer systems worldwide, several studies have observed a significant positive correlation between groundwater O_2 concentration and depth to water (Datry et al., 2004; Foulquier et al., 2010; Goldscheider et al., 2006; McMahon and Chapelle, 2008; Pabich et al., 2001). Where the water table is close to the soil surface, groundwater O_2 consumption is likely rapid because of incomplete degradation of soil-generated labile DOC in the unsaturated zone. On the contrary, where the water table is far from the soil surface, strong oxygen depletion in the vicinity of the water results in almost complete degradation of soil-generated DOC in the unsaturated zone (Malard and Hervant, 1999; Starr and Gillham, 1993). A regional study in forest soils of Switzerland (Hagerdon et al., 2000) and a study compiling a global database of soil carbon (Camino-Serrano et al., 2014) both found that soil-generated DOC was preferentially mobilized under reducing conditions

in soils because of dissolution of Fe oxides. Deeper water tables in croplands do not reach topsoil that exhibits high labile organic C content. Thus, reducing conditions in topsoil and the leaching of soilgenerated DOC are prevented as is the consumption of the groundwater O₂ stock, as occurs in forests during high flow stages (Table 3; Fig. S2l; Deirmendjian et al., 2018). Therefore, groundwater pCO₂ was higher in the forest during high flow than it was in cropland and riparian sites (Table 3; Fig. 4h, S2l, h). This also explains the negative correlation between DOC and CO₂ observed only in forest groundwater (Fig. 6b). During base flow, we observed a clear land use spatial pattern among cropland, riparian forest, and forest sites (Table 3; Fig. S2h). We hypothesize that this difference was a consequence of a less intense soil respiration in croplands during summer. From simultaneous eddy covariance measurements over pine forests and maize croplands of the study area, Stella et al. (2009) confirmed that ecosystem respiration was lower in croplands than in forests over the whole year. However, Stella et al. (2009) also showed that ecosystem respiration was larger during the growing season of the maize, because of increased of soil respiration in response to the higher soil water content caused by irrigation. In forest sites, groundwater pCO_2 increases during the summer because soil CO_2 diffuses downward and then is dissolved into the water table (Deirmendjian et al., 2018; Tsypin and Macpherson, 2012). A deeper water table in cropland suggests a less efficient CO₂ transfer from soil air to water table. Higher soil moisture in croplands due to irrigation probably delays soil CO₂ diffusion to groundwater.

The δ^{13} C-DIC signature of forest groundwater was typical of a signature that originated from respiration of soil organic matter derived from C₃ plants (O'Leary, 1988; Vogel et al., 1993). The studied forest soils have no natural carbonate minerals (Augusto et al., 2010) and DIC originating from silicate weathering has the same isotopic signature as DIC originating from soil respiration (Das et al., 2005; Polsenaere and Abril, 2012; Wachniew, 2006). Crop groundwater had a heavier δ^{13} C-DIC signature than forest groundwater and this discrepancy resulted from distinct processes. Liming in cropland brings artificial carbonates into crop soil and DIC originating from carbonate weathering produced DIC with a δ^{13} C value of approximately half of that of soil CO₂ as carbonate rocks have a δ^{13} C of approximately 0%, making δ^{13} C-DIC less negative (Clark and Fritz, 1997; Salomons and Mook, 1986). Irrigation with extraction of groundwater could also increase the δ^{13} C-DIC signature by enhancing the degassing rate of ¹²CO₂ relative to ¹³CO₂ (Deirmendjian and Abril, 2018; Polsenaere and Abril, 2012). Changes in the δ^{13} C-DIC signature could also originate from respiration of soil organic matter derived from maize, a C₄ plant with a heavier δ^{13} C signature than C₃ forest plants (O'Leary, 1988; Vogel et al., 1993), as observed in the study region (Quénéa et al., 2006). Indeed, after three decades of cultivation, the remaining carbon from the forest pool was mostly recalcitrant and its degradation probably did not affect the δ^{13} C-DIC pool (Jolivet et al., 1997).

4.3. Dynamics of IN and CH₄ in groundwater: The influence of groundwater O_2

Subsurface and groundwater redox zonation is driven by the spatial and temporal distribution of O₂ that serves as the primary terminal electron acceptor during the degradation of organic C. In crop groundwater, high O₂ concentrations inhibited methanogenesis, as this process is strictly anaerobic and thus resulted in very low CH₄ concentrations (Table 2; Fig. 4f, g; Borges et al., 2018; Ciais et al., 2010; Jurado et al., 2017; Klüber and Conrad, 1998). Conversely, forest and riparian groundwater exhibited anoxic conditions that allowed methanogenesis to occur and created higher CH₄ concentration in forest sites compared to cropland sites (Table 2; Fig. 4f, g). High water table stages in forested areas cause anoxia in soils, forcing plants and microorganisms to switch to anaerobic metabolism (Naumburg et al., 2005; Bakker et al., 2006, 2009). Thus, in riparian and forest areas, we expected a positive relationship between water table and groundwater CH₄ but, to the contrary, we observed a negative relationship between these two parameters ($R^2 = 0.25$, p < 0.05; data not shown). This implies that methanogenesis primarily occurs in deeper layers of forest soils, especially in summer. Fe²⁺ and NH₄⁺ accumulates in forest groundwater because anoxic conditions inhibit nitrification and iron oxidation (Table 2; Fig. 4d, e; Jambert et al., 1994; Widdel et al., 1993).

In groundwater, anoxic conditions enable heterotrophic denitrification, whereas an O₂ threshold of 30–60 μ mol L⁻¹ completely inhibits heterotrophic denitrification (Balestrini et al., 2016; Cey et al., 1999; Christensen et al., 2013; Jambert et al., 1994; Kolbjørn Jensen et al., 2017; Korom, 1992). In strictly forest sites, denitrification in groundwater is usually limited by the scarcity of NO_3^- , whereas in strictly crop sites denitrification is often limited by organic C availability (Table 2; Jambert et al., 1994; Starr and Gillham, 1993). Thus, N fertilizer application associated with different groundwater denitrification rates in the different plots creates the observed spatial pattern of groundwater NO_3^- concentration in crop, riparian and forest sites (Table 2; Fig. 4c). In local maize croplands, Jambert et al. (1997) found that 13% of the N fertilizers inputs were converted to N₂ gas, demonstrating that denitrification could occur in these oxic crop soils and groundwater. Although oxic conditions are not favorable for groundwater denitrification, some studies in agricultural catchments do describe this process at relatively high O_2 (150 µmol L⁻¹) levels (McAleer et al., 2017; Otero et al., 2009). In crop soils, Rubol et al. (2016) investigated the spatiotemporal dynamics in oxidative microbial activity and the development of anoxic micro zones (i.e., anoxic hot-spots) at the microscopic level (μ m to cm). They found that labile C addition resulted in maximum rates of local metabolic activity within a few minutes and led to the subsequent formation of anoxic hotspots and thus, both oxic and anoxic conditions coexisted closely within a small volume of crop soils. Consequently, denitrification probably occurs in anoxic microsites in waterlogged soil during irrigation as higher soil moisture results in lower soil oxygen concentration, lower redox potential and higher leaching of soil DOC (Hagedorn et al., 2000; Jambert et al., 1997; Rubol et al., 2012; Silver et al., 1999).

N fertilizer load in local croplands is 25 g N m⁻² yr⁻¹ (Jambert et al., 1997), whereas export (using drainage of 637 mm yr^{-1} and the average NO₃⁻ concentration in crop groundwater) of NO₃⁻ through crop groundwater was estimated at $10 \text{ g N m}^{-2} \text{ yr}^{-1}$ (40% of the annual N fertilizer load), and export (using same drainage and the average NO_3^- concentration in riparian groundwater) of NO₃⁻ through riparian groundwater was estimated at 2.8 g N m⁻² yr⁻¹ (8% of the annual N fertilizer load). This shows the importance of riparian groundwater to attenuate N inputs from adjacent croplands to streams, otherwise a large portion of the annual N fertilizer load would have been leached into adjacent streams rather than being denitrified or used by plants. In riparian groundwater adjacent to a farm in the New York state (USA), Anderson et al. (2014) found that total groundwater denitrification was equivalent to 32% of manure N spread on the adjacent upland field. Mekala et al. (2017) simulated the transport and dynamics of N in an agricultural soil under flooded conditions and concluded that relatively shallow aquifers with sandy soil are vulnerable to NO₃⁻ contamination at around 10 days if continuous irrigation is practiced. They also stated that NO_3^- had higher leaching potential than NH_4^+ or DOC. At our study site, irrigation and associated desorption of DOC and NO₃⁻ could explain their slight increase in crop groundwater during base flow (Table 3, Fig. 1, S2c). In a storm infiltration basin in Florida (USA), O'Reilly et al. (2012) found that concomitant peaks in groundwater O_2 and NO₃ concentrations after storm rainfall were a consequence of organic N leaching, indicating that there were short periods of ammonification and nitrification. In crop groundwater of Wallonia (Belgium), when groundwater O_2 levels are higher than 125 µmol L⁻¹ (as at the study site), nitrification rather than denitrification promotes the accumulation of N₂O in groundwater (Jurado et al., 2017).

4.4. Stream biogeochemical functioning: Mostly a function of groundwater composition

 NO_3^- inputs to streams cause stream eutrophication (Carpenter et al., 1998; Jordan and Weller, 1996; Smith, 2003; Zhou et al., 2017). This is consistent with our field observations where we observed that crop streams were highly vegetated with macrophytes during base flow stages. Compared to high flow conditions, crop stream eutrophication was accompanied by higher pH and δ^{13} C-DIC, and lower pCO₂ caused by preferential ¹²CO₂ uptake during the macrophyte plant photosynthesis (Table 4; Fig. S3a, h, j; De Carvalho et al., 2009; Raven et al., 2002). The development of macrophytes in crop streams modifies flow and can cause a significant drop in water velocity, which in turn, gives rise to extensive deposition and retention of sediment beneath the macrophytes (Cotton et al., 2006; Sand-Jensen and Pedersen, 1999). This leads to seasonal accumulation of organic matter, a predominance of anoxic conditions in stream sediments, and thus the occurrence of methanogenesis as evidenced by peaks in dissolved CH₄ during base flow (Table 4; Fig. S3g; Borges et al., 2018; Crawford et al., 2016; Sanders et al., 2007). Crop stream CH₄ concentration was 390 nmol L^{-1} during base flow (Table 2), a concentration significantly lower (1430 nmol L^{-1}) than chalk streams impacted by macrophyte vegetation in England (Sanders et al., 2007). This discrepancy probably resulted from the increased in silt and clay fraction during summer of the underlying sediment in chalk streams (Sanders et al., 2007). This would suggest that the permeability of chalk stream sediment became

lower than that of sandy stream sediment and created stronger reducing conditions in chalk stream sediments, which likely increased the potential for methanogenesis (Baker et al., 1999; Findlay, 1995; Kankaala et al., 2005; Morrice et al., 1997). Sanders et al. (2007) also showed that the chalk streams' emissions of CH₄ to the atmosphere were approximately 50 times lower than the CH₄ production in stream sediments, illustrating the high potential for CH₄ oxidation in the water column of crop stream. During base flow, a second explanation for higher CH_4 (and NH_4^+) concentrations in crop streams relative to forest streams could be differential hydrology. Drainage plot is a function of the water table height (hydraulic gradient, Darcy's law). During base flow, the water table in cropland was deeper than in forest (e.g., 4 m deeper in Sep. 2014; Fig. 2), and so during this period, potentially more forest groundwater was drained into crop streams. However, in forest streams we usually did not observe CH_4 (or NH_4^+) concentrations as high as in crop streams (Table 2; Fig. 7d) indicating that CH₄ (or NH_{4}^{+}) in crop streams primarily originated from crop stream sediments rather than from higher discharge of forest groundwater. In crop streams, CH_4 was correlated with NH_4^+ but not correlated with $NO_3^$ or DOC (Figs. 6d, 7c, d). Such relationships were also observed in the Meuse river basin (Belgium) (Borges et al., 2018) and in a global meta-analysis of riverine CH₄ (Stanley et al., 2016). In contrast, this does not fit the conceptual model of Schade et al. (2016) developed from data in New Hampshire streams, whereby the CH₄ was positively correlated with DOC, while negatively related to NO₃⁻.

Sandy sediments of low order stream beds impacted by eutrophication are significant areas of NO₃⁻ reduction over the spring and summer, lowering DOC and NO₃⁻ concentrations in stream water (Table 4; Fig. S3c, l; Böhlke et al., 2009; Mulholland et al., 2008; Sanders et al., 2007). Additionally, the decreased of stream velocity during base flow increased residence times of NO_3^- in the hyporheic zone and the time for denitrification (Bardini et al., 2012). In a small stream dominated by maize cropland in the USA, Böhlke et al. (2009) demonstrated that denitrification mainly occurred in sediments and not in the water column since integrated rates of pore-water denitrification derived from ¹⁵N tracer profiles within the hyporheic zone were similar to the reach-scale rates derived from measurements in the stream. In crop streams, a portion of the NO₃⁻ variability between the two hydrological periods could also result from higher drainage of forest groundwater during base flow, which would dilute the NO_3^- signal from crop groundwater.

Considering the flat catchment topography, a minor portion of TSM and POC in streams originates from soil erosion and surface runoff. The most frequent effects of dredging on aquatic ecosystems are changes in the concentration of suspended solids, turbidity and light penetration (Lewis et al., 2001; Newell et al., 1998). Higher concentrations of POC (and TSM) observed in crop streams were also caused by macrophyte biomass developed in summer became a sediment trap. When stream discharge was sufficiently energetic, it re-suspended all the accumulated sediment and removed this litter. Moreover, we observed peaks of CH_4 and NH_4^+ in crop streams during high flow (Table 4; Fig. S3d, g), suggesting that dredging or streambed erosion of crop streams also release CH_4 and NH_4^+ from the sediment.

In forest streams, we observed significantly lower concentrations of Fe^{2+} , NH_4^+ , and CH_4 than in forest groundwater and significant negative linear relationships between O_2 on the one side and Fe^{2+} , NH_4^+ , or CH_4 (Table 2; Figs. 4d, e, g, 7d, f). This suggests there were low O_2 concentration groundwater inputs with high concentrations of reduced compounds and that the stream water was gradually oxygenated, which induced Fe^{2+} and CH_4 oxidations and nitrification. Mulholland et al. (2000) studied N cycling by adding ¹⁵N-labeled NH_4^+ into a forest stream in eastern Tennessee (USA). They concluded that the residence time of NH_4^+ in the water column was low (5 min) and that nitrification was an important sink for NH_4^+ , accounting for 19% of total ammonium uptake. In forest streams, the NH_4^+ concentration was approximately 3 µmol L^{-1} lower than in forest groundwater and thus did not explained

the NO₃⁻ increase of 50 µmol L⁻¹ (Table 2; Fig. 3c, d). Up to 76% of N exports from local forest are in organic forms but these N exports are very low (<0.2 g N m⁻² yr⁻¹; De Wit et al., 2005; Rimmelin, 1998; Vernier et al., 2003), so in-stream mineralization of organic N coupled to nitrification could not explain NO₃⁻ concentrations in forest streams. Since the sampled forest streams are not strictly forested, NO₃⁻ concentration are explained by simple hydrological mixing between crop and forest groundwater (Table 2).

In streams, pCO_2 was lower and O_2 was higher than in groundwater (Table 2; Figs. 3f, h, 6a). This shows that gas exchange between stream water and the atmosphere occurs quickly, favored by low stream depth and strong concentration gradients between the two compartments. Some authors (e.g., Bodmer et al., 2016; Borges et al., 2018) found elevated pCO₂ in crop streams rather than in forest streams, due to higher levels of dissolved and particulate organic matter in crop dominated systems compared to the forested ones that facilitated the in-stream degradation of organic matter. Moreover, land uses are expected to change the composition of terrestrial soil organic matter leached to streams, shifting from vegetation- to microbe-derived organic matter with greater agricultural land use and potentially higher emissions in crop streams (Fuss et al., 2017; Graeber et al., 2015; Wilson and Xenopoulos, 2009). Those results contrasted with ours because we found no difference in pCO₂ between crop and forest streams. Forest groundwater did have higher pCO₂ than crop groundwater, indicating a more intense degassing in forest streams. The similar δ^{13} C-DIC signatures in forest and crop streams despite the strong difference between crop and forest groundwater suggests faster isotopic equilibration of DIC resulting from degassing. The greater gas transfer velocity in forest streams is a consequence of the abundance of coarse woody debris which generates higher levels of water turbulence (e.g., Bodmer et al., 2016), and is consistent with our field observations. A lower gas transfer velocity lower in crop streams results from stream calibration reducing turbulent flow, and macrophyte vegetation that protects the water surface from wind shear.

5. Conclusion

The present study demonstrates that C and IN concentrations in shallow groundwater and in first-order streams are strongly sensitive to land use. In sandy lowland catchments, simultaneous measurements of biogeochemical parameters in groundwater and streams are crucial for identifying and quantifying biogeochemical processes involved at the groundwater-stream interface. We also show that a statistical clustering analysis based on NO_3^- dataset enables partitioning of groundwater and streams into crop-affected or forest-dominated waters. Such a classification could be useful to river managers and policy makers. The water table had greater depth in croplands and was a crucial parameter necessary for understanding groundwater biogeochemical variability in relation to land use. Higher water table stages in forests created anoxic conditions and increased soil leaching. Conversely, in croplands, the deeper water table prevented anoxic conditions, creating different groundwater compositions from forest groundwater and inhibiting the denitrification of the N fertilizers, which led to groundwater $NO_3^$ accumulation. Despite the occurrence of groundwater denitrification in riparian and forest sites, N fertilizers inputs in crop streams were still high enough to generate eutrophic conditions in these streams. Eutrophication resulted in a biogeochemical cascading effect, which sustained high CH₄ concentration and lowered NO₃⁻. High CO₂ and CH₄ production occurs in forest soils and groundwater, but these two gases exhibit lower concentrations in forest streams, indicating intense degassing or oxidation.

The groundwater-stream interface is a biogeochemical hotspot and hot moment for C emissions and N removal processes (McClain et al., 2003). Future studies focusing on the groundwater-stream interface in relation to land use are needed to better understand C and N dynamics in aquatic systems in order to correctly close C and N budgets at regional and global scales.

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Appendix A. Supplementary data

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